

CORONASTEP Report 57 (Week 11) SARS-CoV-2 Sewage Surveillance in Luxembourg

Summary

This report 57 presents the results of SARS-CoV-2 contamination of wastewater at the entrance of the 13 wastewater treatment plants during the week 11 of 2021.

The flux of SARS-CoV-2 RNA present in wastewater treatment plants during the week 11 indicates a still high prevalence of the virus in wastewater at the national level, with a constant trend since the beginning of last week. Next week's sampling will confirm or not the observed dynamics.

The evolution of the SARS-CoV-2 contamination appears also stabilized at the level of the wastewater treatment plant studied individually.

Table 1 – National level of SARS-CoV-2 contamination of wastewaters in Luxembourg.



Dark green: negative samples for SARS-CoV-2 gene E (-), Green to red: positive samples for SARS-CoV-2 gene E. The intensity of the color is related to the national SARS-CoV-2 flux (RNA copies / day / 100 000 equivalent inhabitants).

Contamination Week 3 Level Week 33 Week 11 Week 14 Week 12 Week 15 Week 15 Week 16 Week 16 Week 17 Week 21 Week 22 Week 22 Week 23 Week 24 Week 25 Week 27 Week 26 Week 27 Week 26 Week 27 Week 27 Week 28 Week 31 Week 31 Week 32 Week 32 Week 33 Week 33 Week 34 Week 34 Week 35 Week 35 Week 36 Week 36 Week 37 Week 37 Week 40 Week 41 Week 41 Week 42 Week 41 Week 41 Week 42 Week 42 Week 44 Week 41 Week 42 Week 42 Week 42 Week 42 Week 42 Week 42 Week 42 Week 43 Week 42 Week 44	•	
4. 4. 4. 4. 4. 4. 4. 4. 4. 4. 4. 4. 4. 4	Nationa ntaminat Level	Week
4 + 4 + 4 + 4 + 4 + 4 + 4 + 4 + 4 + 4 +		Week 3
45 4 4 4 4 4 4 5 3 3 3 3 3 3 3 3 3 3 3 3		Week 7
111 111 111 111 111 111 111 111 111 11		Week 9
14		Week 11
110 110 110 110 110 110 110 110 110 110		Week 14
10 10 10 10 10 10 10 10 10 10 10 10 10 1		Week 15
117 118 118 119 119 119 119 119 119 119 119		Week 16
118 119 119 119 119 119 119 119 119 119		Week 17
119 20 20 20 20 22 22 22 22 22 22 22 22 23 33 33 33 33		Week 18
20 21 22 22 22 23 23 23 23 23 23 23		Week 19
221 222 223 224 224 224 226 226 226 227 227 228 233 333 333 333 334 40 40 40 40 40 40 40 40 40 40 40 40 40		Week 20
222 23 23 23 25 25 25 25 25 25 25 25 27 27 27 27 27 27 27 27 27 27 27 27 27		Week 21
23 24 27 27 27 27 27 27 27 27 27 27		Week 22
225 226 226 227 227 227 228 228 238 333 333 334 440 440 440 440 440 440 440		Week 23
255 266 277 277 278 288 383 331 331 331 331 341 341 341 34		Week 24
25 27 27 27 28 33 33 33 33 33 33 33 33 33 33 33 33 33		Week 25
27 28 29 30 30 30 33 31 31 31 32 34 44 44 44 44 44 44 44 44 44 44 44 44		Week 26
288 30 30 33 33 33 34 40 40 40 44 44 44 44 44 44 44 44 44 44		Week 27
29 30 33 31 33 33 33 33 33 34 40 40 40 40 40 40 40 40 40 40 40 40 40		Week 28
30 31 31 33 33 33 33 33 33 34 44 44 44 44 44 44		Week 29
33 33 33 33 33 33 33 33 33 34 40 40 40 40 40 40 40 40 40 40 40 40 40		Week 30
33 33 33 34 35 35 36 36 37 47 47 47 47 47 47 47 47 47 47 47 47 47		Week 31
333 34 34 35 35 36 36 36 40 40 40 41 41 41 41 41 41 41 41 41 41 41 41 41		Week 32
34 35 36 38 38 38 39 40 40 44 44 44 44 44 44 44 44 44 44 44		Week 33
35 36 37 37 39 39 40 40 44 44 44 44 44 45 45		Week 34
36 37 37 39 39 39 40 44 44 44 44 44 44 44 44 45 45		Week 35
37 38 38 39 39 40 41 41 44 44 44 44 44 44 44 44 44 44 44		Week 36
38 39 40 41 42 43 44 44 44 45		Week 37
39 40 41 42 43 44- 44- 45-		Week 38
40 42 43 44- 44- 45-		Week 39
41 42 43 44- 44- 45- 45-		Week 40
42 44- 44- 45- 45-		Week 41
43 44- 44- 45-		Week 42
44- 45- 45-		Week 43
44- 45- 45-		44-
45- 45-		44-
45-		45-
		45-

National Contamination	Week
Level	
	Week 45-3
	Week 46-1
	Week 46-2
	Week 46-3
	Week 47-1
	Week 47-2
	Week 48-1
	Week 48-2
	Week 48-3
	Week 49-1
	Week 49-2
	Week 50-1
	Week 51-2
	Week 52
	Week 53
	Week 01-1
	Week 01-2
	Week 02-1
	Week 02-2
	Week 03-1
	Week 03-2
	Week 04-2
	Week 05-1
	Week 06-1
	90
	Week 07-1
	Week 07-2
	Week 08-1
	Week 08-2
	Week 09-2
	Week 10-2
	11-
	Week 11-2



Figure 1a – RT-qPCR quantification time-course monitoring of SARS-CoV-2 (E gene) in Luxembourgish wastewater samples from December 2019 to March 2021. Grey squares: daily-confirmed cases for Luxembourgish residents (https://data.public.lu/fr/datasets/donnees-covid19/), Blue dots: cumulative SARS-CoV-2 flux (RNA copies / day / 100 000 equivalent inhabitants).

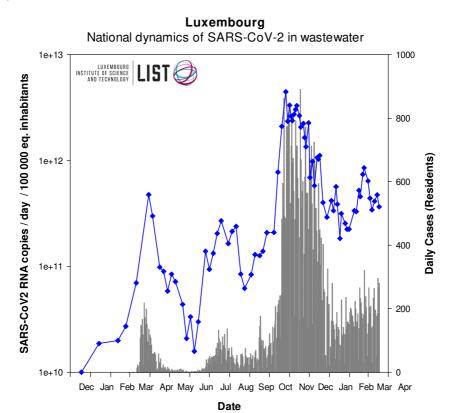


Figure 1b – Close-up of Figure 1a showing results from September 1st on.

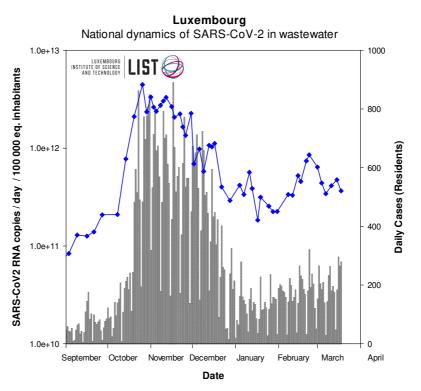




Table 2 - Level of SARS-CoV-2 contamination of each analyzed wastewater treatment plant in Luxembourg during the second wave. BEG: Beggen, BET: Bettembourg, SCH: Schifflange, BLE: Bleesbruck, MER: Mersch, PET: Pétange, HES: Hespérange, ECG: Echternach, UEB: Uebersyren, GRE: Grevenmacher, TRO: Troisvierges, BOE: Boevange sur Attert. WIL: Wiltz



Dark green: negative samples for SARS-CoV-2 gene E (-), Green to red: positive samples for SARS-CoV-2 gene E. The intensity of the color is related to the RT-qPCR signal (Ct values) Grey boxes: no data

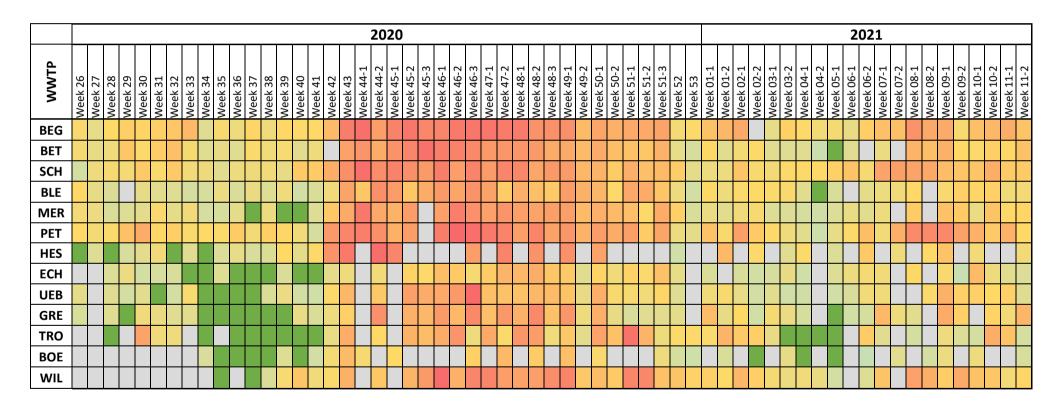




Figure 2a – RT-qPCR quantification time-course monitoring of SARS-CoV-2 (E gene) in the four most impacted wastewater treatment plants from March 2020 to March 2021. Grey squares: daily-confirmed cases for the contributory area of each wastewater treatment plant, dots: SARS-CoV-2 flux (RNA copies / day / 10 000 equivalent inhabitants).

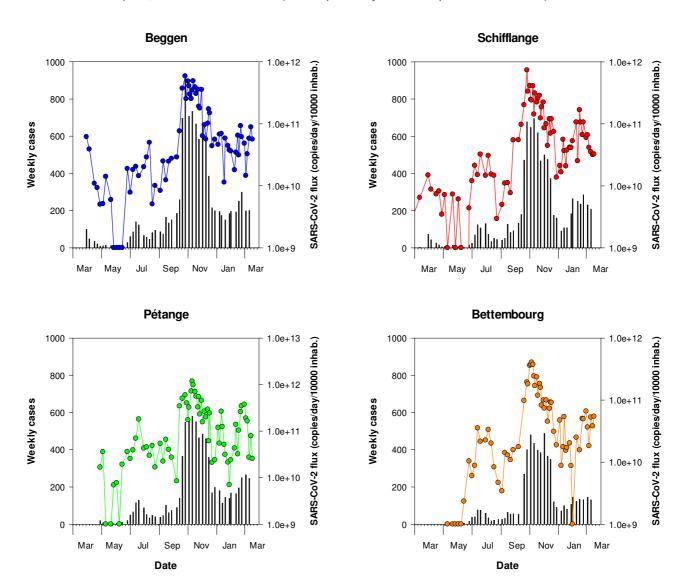


Figure 2b – Close-up of Figure 2a showing results from September 1st on.

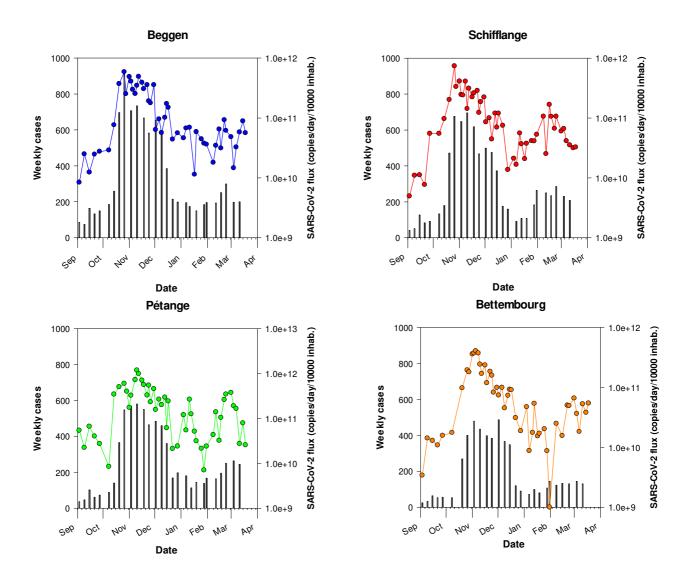
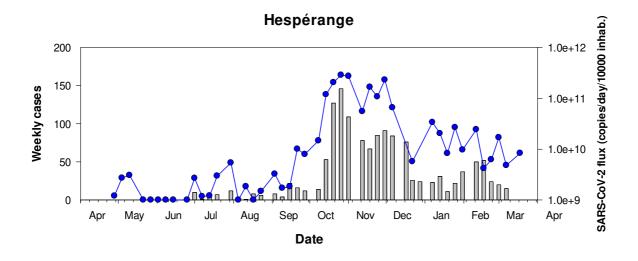
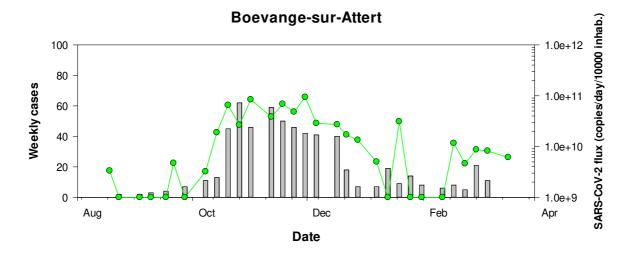




Figure 3a – RT-qPCR quantification time-course monitoring of SARS-CoV-2 (E gene) in Hespérange, Mersch and Boevange-sur-Attert wastewater treatment plants from March 2020 to March 2021. Grey squares: daily-confirmed cases for the contributory area of each wastewater treatment plant, dots: SARS-CoV-2 flux (RNA copies / day / 10 000 equivalent inhabitants).





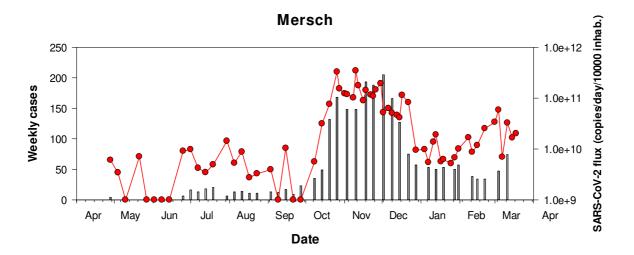


Figure 3b – Close-up of Figure 3a showing results from September 1st on.

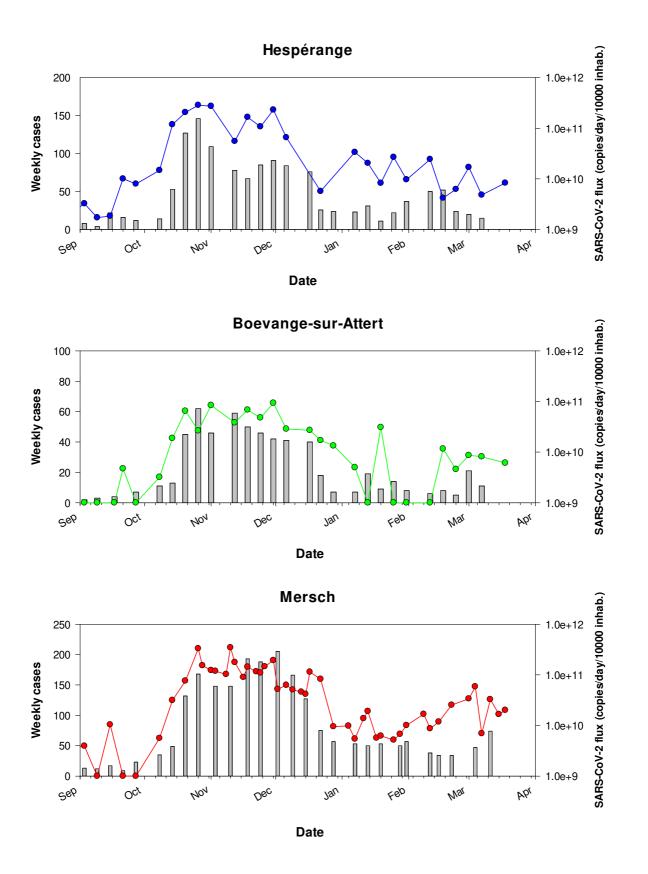
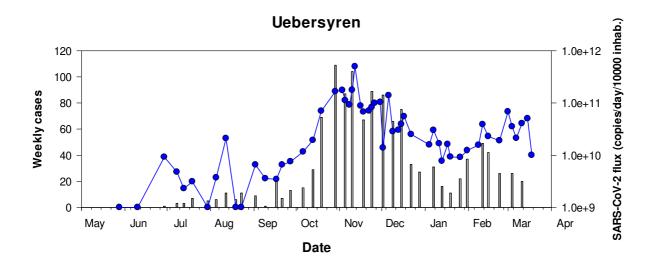
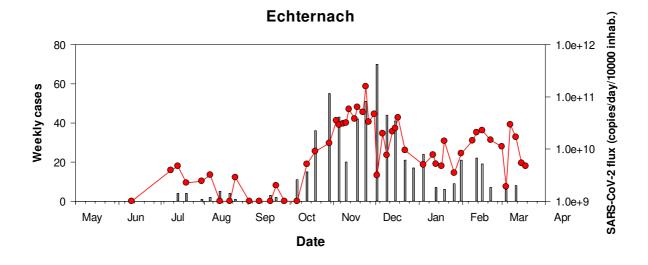




Figure 4a – RT-qPCR quantification time-course monitoring of SARS-CoV-2 (E gene) in SIDEST wastewater treatment plants from March 2020 to March 2021. Grey squares: daily-confirmed cases for the contributory area of each wastewater treatment plant, dots: SARS-CoV-2 flux (RNA copies / day / 10 000 equivalent inhabitants).





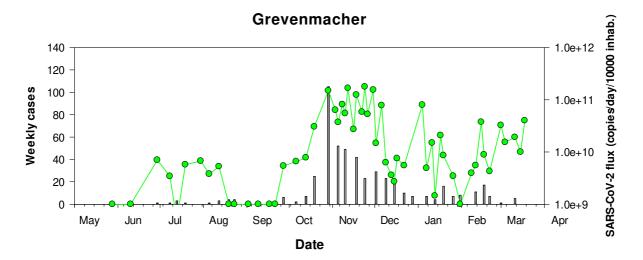
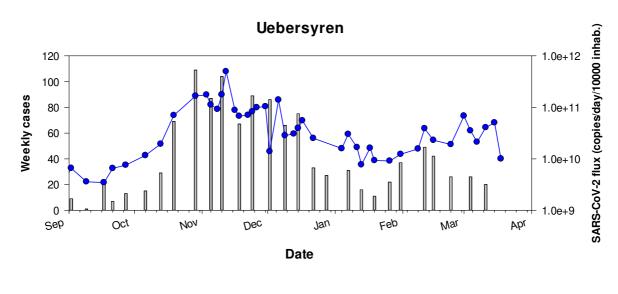
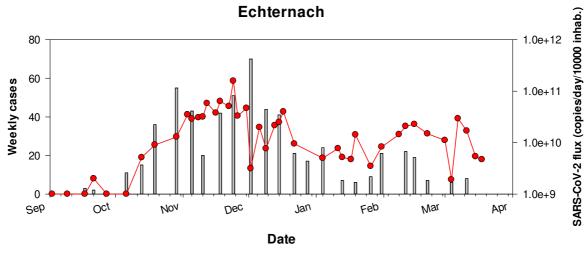


Figure 4b - Close-up of Figure 4a showing results from September 1st on.





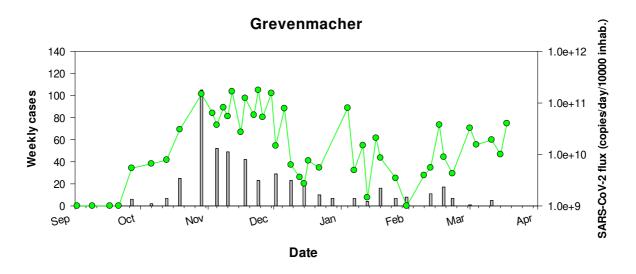
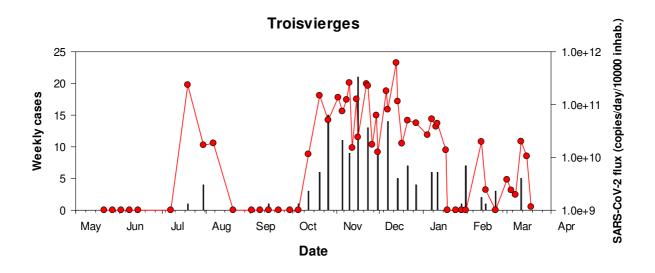
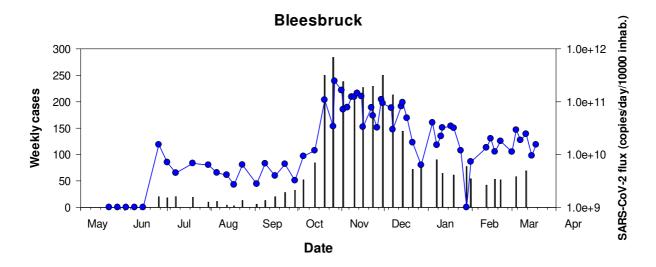




Figure 5a – RT-qPCR quantification time-course monitoring of SARS-CoV-2 (E gene) in SIDEN wastewater treatment plants from March 2020 to March 2021. Grey squares: daily-confirmed cases for the contributory area of each wastewater treatment plant, dots: SARS-CoV-2 flux (RNA copies / day / 10 000 equivalent inhabitants).





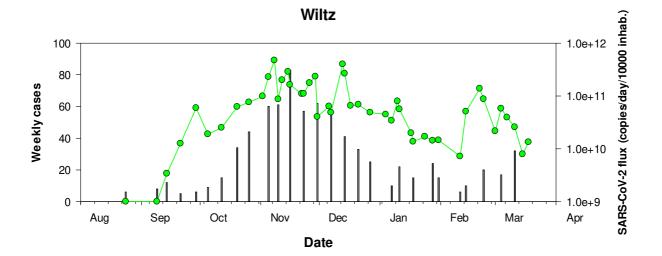


Figure 5b - Close-up of Figure 5a showing results from September 1st on.

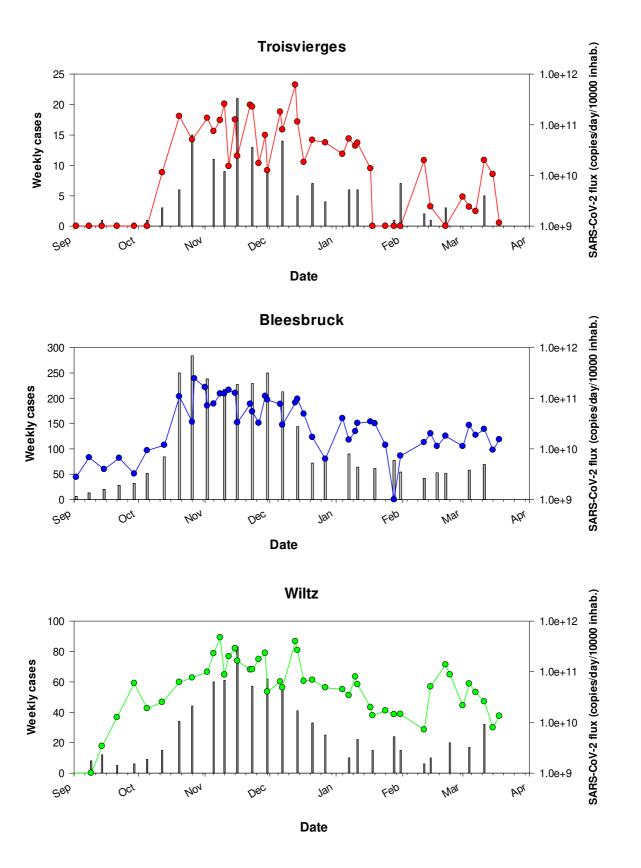


Table 3- Timing of sewage sampling since the beginning of the CORONASTEP study

																						202	0																								20	21					
WWTP	Max capacity (eq. inhabitants)	Inhabitants connected	Week 3	Week 7	Week 11	Week 14	Week 15	Week 16	Week 17	Week 18	Week 19	Week 20	Week 21	Week 23	Week 24	Week 25	Week 26	Week 27	Week 28	Week 29	Week 30	Week 31	Week 32 Week 33	Week 34	Week 35	Week 36	Week 37	Week 38	Week 39	Week 40	Week 41	Week 42	Week 44	Week 45	Week 46	Week 47	Week 48	Week 49	Week 50	Week 51	Week 52	Week 53	Week 01	Week 02	Week 03	Week 05	Week 05	Week 00	Week 08	Week 09	Week 10	Week 11	Total samples
Beggen	210000	139731				1	1	1	1	1	1	1	1 :	1 1	1	1	1	1	1	1	1	1	1 1	1	1	1	1	1	1	1 :	1 :	1 1	L 2	3	3	2	3	2	2	3	1	1	2	1	2 2	2 2	2 2	2 2	2	2	2	2	73
Bettembourg	95000	53606										1	1 1	1 1	1	1	1	1	1	1	1	1	1 1	1	1	1	1	1	1	1 :	1	1	L 2	3	3	2	3	2	2	3	1	1	2	2	2 2	2 2	2 1	1 2	2	2	2		66
Schifflange	90000	68143	1	1 1	1	1	1	1	1	1	1	1	1 1	1 1	1	1	1	1	1	1	1	1	1 1	1	1	1	1	1	1	1 :	1 :	1 1	L 2	3	3	2	3	2	2	3	1	1	2	2	2 2	2 2	2 2	2 2	2	2	2	2	82
Bleesbrück	80000	30930											1 1	1 1	1	1	1	1	1		1	1	1 1	1	1	1	1	1	1	1 :	1 :	1 1	L 2	3	3	2	3	2	2	3	1	1	2	2	2 2	2 2	2 1	1 2	2	2	2	2	65
Mersch	70000	30473								1	1	1	1 :	1 1	1	1	1	1	1	1	1	1	1 1	1	1	1	1	1	1	1 :	1 :	1 1	L 2	2	3	2	3	2	2	3	1	1	2	2	2 2	2 2	2 2	2 2	2	2	2		69
Pétange	50000	59481	1	1 :	1					1	1	1	1 1	1 1	1	1	1	1	1	1	1	1	1 1	1	1	1	1	1	1	1	1 :	1 1	L 2	2	3	2	3	2	2	3	1	1	2	2	2 2	2 2	2 2	2 2	2	2	2	2	77
Hespérange	36000	15479								1	1	1	1 1	1 1	1	1	1	1	1	1	1	1	1 1	1	1	1	1	1	1	1	1 :	1 1	1	1	1	1	2	1	1	1	1	0	1	1	1 :	1	1	l 1	1	1	1	1	47
Echternach	36000	7499													1				1	1	1	1	1 1	1	1	1	1	1	1	1 :	1 :	1 1	1	2	3	2	3	2	2	3	1	0	1	2	2 :	1 2	2 2	2 2	2	2	2		56
Uebersyren	35000	18600											1	1	1		1		1	1	1	1	1 1	1	1	1	1	1	1	1	1 :	1 1	1	2	3	2	3	2	2	3	1	0	2	2	2 :	1 2	2 2	2 2	1	2	2	2	58
Grevenmacher	47000	9835											- 1	1	1		1		1	1	1	1	1 1	1	1	1	1	1	1	1 :	1 :	1 1	l 1	2	3	2	3	2	2	3	1	0	2	2	2 :	1 2	2 2	2 2	2	2	1		58
Troisvierges	5000	3411											1 1	1 1	1	1			1		1	1	1	1		1	1	1	1	1 :	1 :	1 1	1	2	3	2	3	2	2	3	1	1	2	2	2 2	2 2	2 1	1 2	2	2	2		59
Boevange sur Attert	15000	1170																						1	1	1	1	1	1	1	1 :	1 1	1	1	1	1	2	1	1	1	1	1	1	1	1 :	1	1	l 1	1	1	1		32
Wiltz	16500	6944																							1		1	1	1	1 :	1 :	1 1	1 1	2	3	2	3	2	2	3	1	1	2	2	2 2	2 2	2 1	1 2	2	2	2	2	49
Total	785500	445302	2	2 2	2	2	2	2	2	5	5 (6	8 1	0 8	11	8	9	7	11	9	11	11 :	11 10	12	12	12	13	13	13	13 1	3 1	2 1	3 19	28	35	24	37	24	24	35	13	9	23	23 2	24 2	1 2	4 2	0 24	1 23	24	23	24	791
Pop Lux (2019)		613901																																																			
		72.54%				Т																																								Т	Т			Т			



Materials and Methods

Sewage samples

From March 2020 to March 2021, up to thirteen wastewater treatment plants (WWTPs) were sampled at their inlet according to the planning presented in Table 3. The operators of the WWTPs collected a 24-h composite sample according to their routine sampling procedure. Composite sample was stored at 4°C until sample processing.

Sample processing

The samples were transported to the laboratory at 4°C and viral RNA was isolated on the day of sampling. Larger particles (debris, bacteria) were removed from the samples by centrifugation at 2,400 x g for 20 min at 4°C. A volume of 120 mL of supernatant was filtered through Amicon® Plus-15 centrifugal ultrafilter with a cut-off of 10 kDa (Millipore) by centrifugation at 3,220 x g for 25 min at 4°C. The resulting concentrate was collected and 140 μ L of each concentrate was then processed to extract viral RNA using the QIAamp Viral RNA mini kit (Qiagen) according to the manufacturer's protocol. Elution of RNA was done in 60 μ L of elution buffer.

Real-time One-Step RT-PCR

Samples were screened for the presence of *Sarbecovirus* (*Coronaviridae*, *Betacoronaviruses*) and/or SARS-CoV-2 virus RNA by two distinct real-time one-step RT-PCR assays, trageting the E gene (Envelope small membrane protein) and the N gene (nucleoprotein). The E gene real-time RT-PCR can detect *Sarbecoviruses*, i.e. SARS-CoV, SARS-CoV-2 and closely related bat viruses. In the context of the COVID19 pandemic, it can be assumed that only SARS-CoV-2 strains will be detected by this assay given that SARS-CoV virus has been eradicated and other bat viruses do not commonly circulate in the human population. The E gene assay is adapted from Corman et al. [17]. The N gene real-time RT-PCR assay (N1 assay) specifically detects SARS-CoV-2 virus. It is adapted from the CDC protocol¹. The two primers/probe sets are presented in Table 3. The RT-qPCR protocols and reagents were all provided by the LIH.

Table 4 - RT-qPCR primer-probe sets

Target	Primer name	Primer sequence (5' to 3')	References
E gene	E_Sarbeco_F1	5-ACAGGTACGTTAATAGTTAATAGCGT-3	Corman et al.,
	E_Sarbeco_R2	5-ATATTGCAGCAGTACGCACACA-3	2020
	E_Sarbeco_P1	5'-FAM-ACACTAGCCATCCTTACTGCGCTTCG-BHQ1	
N gene	2019-nCoV_N1_Fw	5'-GAC CCC AAA ATC AGC GAA AT-3'	CDC, 2019
	2019-nCoV_N1_Rv	5'-TCT GGT TAC TGC CAG TTG AAT CTG-3'	
	2019-nCoV_N1 Probe	5'-FAM-ACC CCG CAT TAC GTT TGG TGG ACC-BHQ1-3'	

Each reaction contained 5 μ L of RNA template, 5 μ L of TaqPath 1-step RT-qPCR MasterMix (A15299, Life Technologies), 0.5 μ L of each primer (20 μ M) and probe (5 μ M) and the reaction volume was adjusted to a final volume of 20 μ L with molecular biology grade water. Thermal cycling reactions were carried out at 50 °C for 15 min, followed by 95 °C for 2 min and 45 cycles of 95 °C for 3 sec and 58 °C (E gene) or 55 °C (N gene) for 30 sec using a Viia7 Real-Time PCR Detection System (Life Technologies). Reactions were considered positive (limit of detection – LOD) if the cycle threshold (Ct value) was below 40 cycles.

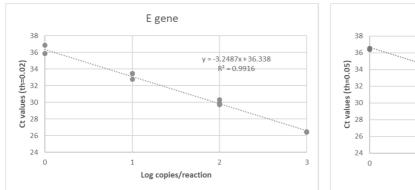
¹ https://www.cdc.gov/coronavirus/2019-ncov/downloads/rt-pcr-panel-primer-probes.pdf



Controls

A non-target RNA fragment commercially available (VetMAX™ Xeno™ IPC and VetMAX™ Xeno™ IPC Assay, ThermoFischer Scientific) was added to the viral RNA extract from sewage concentrates as an internal positive control (IPC). This IPC-RNA is used to control the performance of the RT-qPCR (E gene) and to detect the presence of RT-qPCR inhibitors.

Viral RNA copies quantification of both targeting genes in wastewater samples was performed using RT-qPCR standard curves generated using EDX SARS-CoV-2 Standard (Biorad). This standard is manufactured with synthetic RNA transcripts containing 5 targets (E, N, S, ORF1a, and RdRP genes of SARS-CoV-2, 200,000 copies/mL each). Using such a standard, the limits of quantification (LOQ) of both RT-qPCR assays were estimated to 1 RNA copy per reaction (Figure 6).



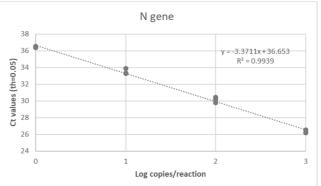


Figure 6 – RT-qPCR standard curves established for both target genes (E gene and N gene) of SARS-CoV-2 using a commercially available standard (Biorad).

Data interpretation

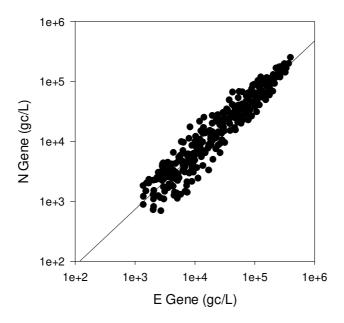
A sample is declared positive for the presence of SARS-CoV-2 if both targets (E and N gene) are detected with Ct values less than or equal to the LOQ. If only one target is detected or if target genes are detected with Ct values between the LOD and the LOQ, samples are reported as presumptive positive (+/-). A sample is declared negative when no target genes are detected (Ct values superior to the LOD).

In case of presumptive positive, sample is tested again using another RT-qPCR detection assay (Allplex 2019-nCoV Assay, Seegene). This commercially available detection kit is a multiplex real-time RT-PCR assay for simultaneous detection of three target genes of SARS-CoV-2 in a single tube. The assay is designed to detect RdRP and N genes specific for SARS-CoV-2, and E gene specific for all *Sarbecovirus* including SARS-CoV-2.

As shown in Figure 7, a highly significant correlation (Pearson Correlation, R^2 =0.964, p = 5.979.10⁻²⁴) was obtained between the SARS-CoV-2 RNA concentrations estimated using the E gene and the N gene, respectively. Therefore, only the E gene results were presented in this report.



Figure 7 - Relationship between the SARS-CoV-2 RNA concentration (RNA copies / L of wastewater) estimated by the both distinct RT-qPCR systems targeting the E and N gene, respectively (n=415),



Acknowledgments

This work is supported by the Fond National de la Recherche (FNR) under project 14806023 - CORONASTEP+ and is conducted in collaboration with the Luxembourg Institute of Health (LIH), the "Laboratoire National de Santé" (LNS) and the University of Luxembourg (LCSB).

In addition, the authors of this report would like to thank all the wastewater syndicates (SIACH, SIVEC, STEP, SIDERO, SIDEN and SIDEST), the "Ville du Luxembourg", the Hespérange city as well as the "Administration de la Gestion de l'Eau" (AGE) for their kind and valuable assistance in the sample collection, the acquisition of wastewater parameters and the collection of demographic data. The authors would also like to thank the Ministry of Heath and the Inspection Sanitaire for their valuable contribution in providing the COVID-19 data at the national and regional scale.